

Note

Blood-group Ii-active oligosaccharides. Synthesis of *O*- β -D-galactopyranosyl-(1 \rightarrow 4)-*O*-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)-D-mannose

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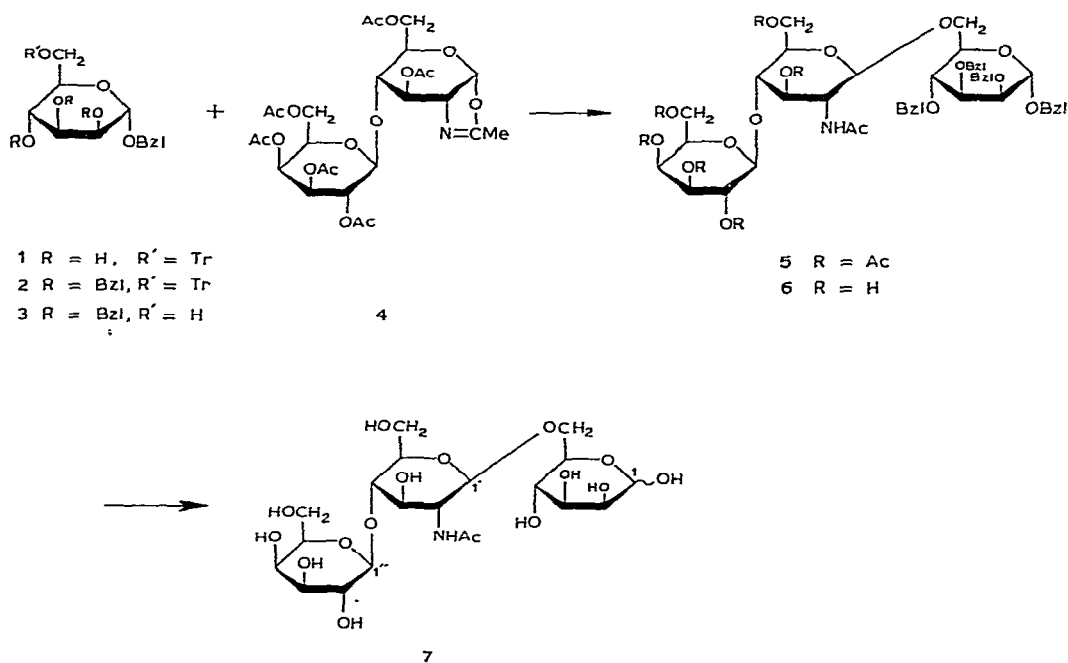
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In a preceding paper¹, we described the synthesis of the trisaccharide *O*- β -D-galactopyranosyl-(1 \rightarrow 4)-*O*-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 3)-D-mannose; this sequence had been observed² in the carbohydrate chain of some glycopeptides isolated from calf-thymocyte, plasma membranes, where two, outer lactosamine-chains are linked to C-3 and C-6 of the same outer D-mannose residue. These structures may be postulated to be present also in the major sialoglycoprotein of human erythrocyte membranes and be responsible for some blood-group Ii activity³. These considerations prompted us to synthesize oligosaccharides that could be used in hemagglutination-inhibition studies. We now report the synthesis of the trisaccharide *O*- β -D-galactopyranosyl-(1 \rightarrow 4)-*O*-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)-D-mannose (7). A similar sequence where the terminal, D-mannose reducing unit is replaced by a D-galactose residue has been shown⁴ to be recognized by two anti-I sera (Ma and Woj).

In 1972, Matta and Bahl⁵ coupled 2-methyl-[3,4,6-tri-*O*-acetyl-1,2-dideoxy- α -D-glucopyrano]-[2,1-*d*]-2-oxazoline with 1,2,3,4-tetra-*O*-acetyl- β -D-mannopyranose in 26% yield. When the condensation of the disaccharide oxazoline, 2-methyl-[3,6-di-*O*-acetyl-1,2-dideoxy-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-galactopyranosyl)- α -D-glucopyrano]-[2,1-*d*]-2-oxazoline⁶ (4) was attempted with the same D-mannopyranosyl derivative, intractable mixtures were always obtained. We observed that prolonged heating at 60° of the D-mannopyranosyl derivative alone, in the presence of catalytic amounts of *p*-toluenesulfonic acid, gave rise to many compounds susceptible of coupling with the glycosylating agent.

An acid-stable derivative of D-mannopyranose was therefore required for condensation with a disaccharide oxazoline. Benzyl 2,3,4-tri-*O*-benzyl- α -D-mannopyranoside (3) was easily prepared by *O*-benzylation of benzyl 6-*O*-trityl- α -D-mannopyranoside¹ (1), followed by acid hydrolysis of the trityl group. Compound 3

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was found to be identical with the compound obtained⁷ by hydrogenolysis of benzyl 2,3-di-*O*-benzyl-4,6-*O*-benzylidene- α -D-mannopyranoside with lithium aluminium-hydride-aluminium chloride.

Condensation of 3 with oxazoline 4 was conducted in 1,2-dichloroethane for 60 h at 50°, in the presence of *p*-toluenesulfonic acid, and gave the protected trisaccharide 5 as a syrup in 52% yield. *O*-Deacetylation afforded 6, crystallized as a hemihydrate. Removal of the *O*-benzyl groups by catalytic hydrogenolysis gave the free trisaccharide 7, obtained as an amorphous solid. Its 250-MHz ¹H-n.m.r. spectrum showed two signals, at δ 5.12 and 4.85, corresponding to the anomeric proton of the reducing D-mannose unit, which exists as two α - and β -D-pyranose configurations in a ratio of $\sim 1:1$. The β -D configuration of the 2-acetamido-2-deoxy-D-glucopyranosyl residue was confirmed by the signal of the anomeric proton (δ 4.56, $J_{1',2'} 7.5$ Hz). The anomeric proton of the terminal D-galactose residue gave a signal at the usual location⁴ (δ 4.45, $J_{1'',2''} 7.5$ Hz).

EXPERIMENTAL

General methods. — Optical rotations were measured at 20° with a Roussel-Jouan electronic, digital micropolarimeter. N.m.r. spectra were recorded at 250 MHz with a Cameca model STN 250 spectrometer, equipped with a Fourier-transform unit for solutions in [²H]chloroform and with tetramethylsilane as internal standard, or in deuterium oxide with tetramethylsilane (0.2% solution in [²H]chloroform) as external reference. T.l.c. was performed on plates of silica gel (with fluorescence

indicator; layer thickness 0.25 mm; E. Merck, Darmstadt, Germany). The compounds were detected by spraying the plates with 1:19 (v/v) conc. sulfuric acid-ethanol. Silica gel Merck (70–325 mesh; E. Merck) was used for column chromatography. Paper chromatography was performed on Whatman No. 1 paper. Free sugars were detected with the aniline hydrogenphthalate reagent. Microanalyses were performed by the Laboratoire Central de Micro-Analyse du C.N.R.S.

Benzyl 2,3,4-tri-O-benzyl-6-O-trityl- α -D-mannopyranoside (2). — A solution of benzyl 6-O-trityl- α -D-mannopyranoside (**1**) (5.12 g, 10 mmol) in *N,N*-dimethylformamide (60 mL) was treated with sodium hydride (1.20 g, 50 mmol) and benzyl bromide (6 mL 50 mmol) for 20 h at room temperature. The excess of hydride was decomposed by the addition of methanol to the ice-cooled mixture. Ether (600 mL) was added, and the solution was washed with water, dried (magnesium sulfate), and evaporated. The residue was chromatographed on silica gel with 1:9 (v/v) ether–light petroleum, to give **2** as a syrup (5.2 g, 67%), $[\alpha]_D^{20} +36^\circ$ (*c* 0.84, chloroform).

Anal. Calc. for $C_{53}H_{50}O_6$: C, 81.30; H, 6.44; O, 12.26. Found: C, 81.13; H, 6.46; O, 12.09.

Benzyl 2,3,4-tri-O-benzyl- α -D-mannopyranoside (3). — A solution of **2** (2.20 g, 2.8 mmol) in acetone (36 mL) and *m* hydrochloric acid (4 mL) was heated under reflux for 2 h. The cooled mixture was neutralized with solid sodium hydrogen-carbonate, and the acetone was evaporated. The residue was extracted with chloroform; the extract was washed with water, dried (magnesium sulfate), and evaporated. The resulting material was chromatographed on silica gel with 1:9 (v/v) ethyl acetate–toluene to give **3** as a syrup (1.14 g, 75%), $[\alpha]_D^{20} +55^\circ$ (*c* 1.48, chloroform); lit.⁶ $[\alpha]_D +54^\circ$ (*c* 0.3, chloroform).

Anal. Calc. for $C_{34}H_{36}O_6$: C, 75.53; H, 6.71; O, 17.76. Found: C, 75.27; H, 6.73; O, 17.92.

Benzyl O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)-(1 \rightarrow 4)-O-(2-acetamido-3,6-di-O-acetyl-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)-2,3,4-tri-O-benzyl- α -D-mannopyranoside (5). — To a solution of **3** (0.727 g, 1.35 mmol) in dry 1,2-dichloroethane (25 mL) were added anhydrous *p*-toluenesulfonic acid (30 mg) and a solution of the oxazoline⁶ **4** (0.850 g, 1.38 mmol) in dry 1,2-dichloroethane (8.5 mL). The mixture was stirred under nitrogen for 60 h at 60°, further addition of **4** (0.850 g, 1.38 mmol) and *p*-toluenesulfonic acid (40 mg) being made after 24 h. T.l.c. in 7:7:1 (v/v) benzene–ether–methanol showed the presence of unchanged alcohol **3** (R_F 0.92), a major new compound (R_F 0.60), and only traces of oxazoline **4** (R_F 0.48). The reaction was stopped at this stage, as many decomposition products arising from the oxazoline had begun to appear. The solution was cooled, diluted with dichloromethane, washed with a saturated solution of sodium hydrogencarbonate and then with water, dried (magnesium sulfate), and evaporated. The residue was chromatographed on silica gel with 1:2 (v/v) ethyl acetate–toluene to give **5** as a syrup (0.805 g, 52%), $[\alpha]_D^{20} +9^\circ$ (*c* 1.51, chloroform); n.m.r.: δ 7.36–7.20 (20 H, 4 Ph), 5.56 (d, 1 H, *J* 9 Hz, NH), 5.36 (d, 1 H, $J_{3'',4''} = J_{4'',5''}$ 3.5 Hz, H-4''), 2.16, 2.06, 2.04, 2.02, and 1.96 (18 H, 6 OAc), and 1.82 (s, 3 H, NAc).

Anal. Calc. for $C_{60}H_{71}NO_{22}$: C, 62.22; H, 6.17; N, 1.21; O, 30.40. Found: C, 62.10; H, 6.30; N, 1.04; O, 29.80.

Benzyl O-β-D-galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→6)-2,3,4-tri-O-benzyl-α-D-mannopyranoside (6). — Compound 5 (570 mg) was *O*-deacetylated overnight at room temperature with 50mm sodium methoxide in methanol (20 mL). Methanol (110 mL) was added to dissolve a white precipitate, and the solution was neutralized with Amberlite IR-120 (H^+) ion-exchange resin, filtered, and evaporated. The residue crystallized from methanol to give 6 (315 mg, 70%), m.p. 210–211°, $[\alpha]_D^{20} + 11^\circ$ (*c* 1.38, chloroform).

Anal. Calc. for $C_{48}H_{59}NO_{16} \cdot 0.5 H_2O$: C, 63.01; H, 6.61; N, 1.53; O, 28.85. Found: C, 62.98; H, 6.55; N, 1.39; O, 29.02.

O-β-D-Galactopyranosyl-(1→4)-O-(2-acetamido-2-deoxy-β-D-glucopyranosyl)-(1→6)-D-mannose (7). — A solution of 6 (490 mg) in glacial acetic acid (40 mL) was hydrogenated catalytically in the presence of 10% palladium-on-charcoal (500 mg) for 3 days at room temperature and atmospheric pressure. The mixture was evaporated to dryness, without removal of the catalyst. The residue was applied to a column of silica gel. Elution with 1:3:3, and then 1:2:2 (v/v) water–ethyl acetate–2-propanol afforded pure trisaccharide 7 as an amorphous solid (175 mg, 50%), $[\alpha]_D^{20} - 12^\circ$ (at equilibrium, *c* 1.72, water). This material showed a single spot in t.l.c. on silica gel with 3:3:2 (v/v) water–ethyl acetate–2-propanol (R_{Glc} 0.46), and in paper chromatography with 2:1:2 (v/v) (upper layer) ethyl acetate–pyridine–water (R_{Glc} 0.50 and $R_{lactose}$ 0.76); n.m.r. (D_2O): δ 5.12 (s, 0.5 H, 1-H β), 4.85 (s, 0.5 H, 1-H α); 4.56 (d, 1 H, $J_{1,2}$ 7.5 Hz, 1'-H), 4.45 (d, 1 H, $J_{1'',2''}$ 7.5 Hz, 1''-H), and 2.03 (s, 3 H, NAc).

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REFERENCES

- 1 J. ALAIS AND A. VEYRIÈRES, *J. Chem. Soc., Perkin Trans. I.*, (1981) 2,377.
- 2 R. KORNFELD, *Biochemistry*, 17 (1978) 1415–1423.
- 3 T. FEIZI, A. KAPADIA, AND W. J. YOUNT, *Proc. Natl. Acad. Sci. U.S.A.*, 77 (1980) 376–380; R. A. CHILDS, T. FEIZI, M. FUKUDA, AND S. HAKOMORI, *Biochem. J.*, 173 (1978) 333–336.
- 4 T. FEIZI, E. WOOD, C. AUGÉ, S. DAVID, AND A. VEYRIÈRES, *Immunochemistry*, 15 (1978) 733–736; C. AUGÉ, S. DAVID, AND A. VEYRIÈRES, *Nouv. J. Chim.*, 3 (1979) 491–497.
- 5 K. L. MATTA AND O. P. BAHL, *Carbohydr. Res.*, 21 (1972) 460–464.
- 6 B. A. DMITRIEV, YU. A. KNIREL, AND N. K. KOCHETKOV, *Izv. Akad. Nauk S.S.S.R., Ser. Khim.*, (1974) 411–416; R. KAIFU AND T. OSAWA, *Carbohydr. Res.*, 52 (1976) 179–185.
- 7 A. LIPTAK, I. JODÁL, AND P. NÁNÁSI, *Carbohydr. Res.*, 44 (1975) 1–11.